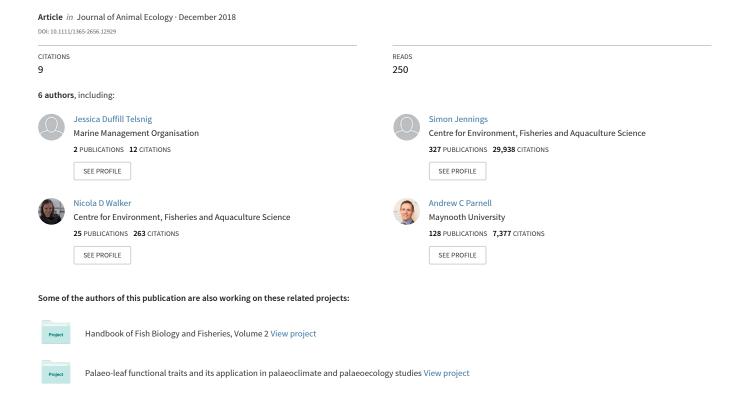
Estimating contributions of pelagic and benthic pathways to consumer production in coupled marine food webs



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Estimating contributions of pelagic and benthic pathways to consumer

production in coupled marine food webs

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Abstract

- Pelagic and benthic systems usually interact, but their dynamics and production rates differ.
 Such differences influence the distribution, reproductive cycles, growth rates, stability and productivity of the consumers they support. Consumer preferences for, and dependence on, pelagic or benthic production are governed by the availability of these sources of production and consumer life history, distribution, habitat, behavioural ecology, ontogenetic stage and morphology.
- 2. Diet studies may demonstrate the extent to which consumers feed on prey in pelagic or benthic environments. But they do not discriminate benthic production directly supported by phytoplankton from benthic production recycled through detrital pathways. The former will track the dynamics of phytoplankton production more closely than the latter.

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- 3. We develop and apply a new analytical method that uses carbon (C) and sulfur (S) natural abundance stable isotope data to assess the relative contribution of pelagic and benthic pathways to fish consumer production.
- 4. For 13 species of fish that dominate community biomass in the northern North Sea (estimated >90% of total biomass), relative modal use of pelagic pathways ranged from <25% to >85%. Use of both C and S isotopes as opposed to just C reduced uncertainty in relative modal use estimates. Temporal comparisons of relative modal use of pelagic and benthic pathways revealed similar ranking of species dependency over four years, but annual variation in relative modal use within species was typically 10-40%.
- 5. For the total fish consumer biomass in the study region, the C and S method linked approximately 70% and 30% of biomass to pelagic and benthic pathways respectively. As well as providing a new method to define consumers' links to pelagic and benthic pathways our results demonstrate that a substantial proportion of fish biomass, and by inference production, in the northern North Sea is supported by production that has passed through transformations on the seabed.

Keywords

Bayesian mixing model, benthic-pelagic coupling, fish, food webs, marine ecosystems, stable isotope analysis

Introduction

In shallow shelf seas, a large proportion of the detritus produced in the water column will accumulate on the seabed and is consumed by seabed (benthic) fauna (Noji, Noji, & Barthel, 1993; Kiørboe, Lundsgaard, Olesen, & Hansen, 1994; Carstensen, Conley, & Müller-Karulis, 2003; Steele & Collie, 2005). Phytoplankton primary production is mostly consumed in the water column, although in shallow tidally mixed regions both seabed fauna and zooplankton can be major phytoplankton consumers (Norén, Haamer, & Lindahl, 1999). In deeper areas more detritus is broken down by microbial activity in the water column and flux to the seabed is relatively low (Suess, 1980; Marsay et al., 2015). Detritus-based production cycles are typically dampened and attenuated in relation to cycles in primary production. The consequence is that continental shelf consumers rely on one or both of two quite distinct trophic pathways, namely those supported directly by primary production or on those supported by detritus (Moore et al., 2004).

Differences in the production dynamics of pelagic and benthic detrital pathways influence the distribution, reproductive cycles, growth rates, stability and productivity of the consumers they support (Hargrave, 1973; Burrel, 1988; Graf, 1992; Marcus & Marcus, 1998; Levinton & Kelaher, 2004). Food web models suggest that the presence of pathways supported by primary production and detritus, dubbed "fast" and "slow" respectively, help to stabilise food webs (McCann, Rasmussen, & Umbanhowar, 2005). Further, stronger coupling between these pathways may increase food web resilience to human and environmental perturbations (Blanchard, Law, Castle, & Jennings, 2011). Various factors may also influence species' affiliation to the pathways such as interannual and consumer-size related variation. For example, changes in planktonic productivity can alter the strength of benthic-pelagic coupling (Woodland & Secor, 2013). Furthermore, small-bodied species and juvenile life stages might depend more on the pelagic pathway due to limited gape size

and higher turnover rates (Woodland & Secor, 2013). Larger individuals with fewer prey size constraints may switch between available prey types, resulting in an increase in trophic reliance on both pathways (Scharf, Juanes, & Rountree, 2000; Woodland & Secor, 2013).

Diet data alone will not discriminate benthic production directly supported by phytoplankton from benthic production recycled through seabed pathways, although diet data can be used as inputs to food web models to achieve this if the diet composition of all interacting consumers is well resolved (Polunin & Pinnegar, 2002). Stable isotope analysis of consumer tissue have been used to measure the extent to which consumers feed on prey from pelagic or benthic environments (Woodland & Secor, 2013; Kopp, Lefebvre, Cachera, Villanueva, & Ernande, 2015) and has the potential to discriminate benthic production directly supported by phytoplankton from benthic production recycled through seabed pathways, with the advantage that stable isotope signatures of larger consumers reflect the composition of assimilated diet over several months (Fry, 2006; Michener & Kaufman, 2007).

Several previous studies of marine pelagic-benthic coupling have shown that consumer $\delta^{13}C$ can be linked to use of benthic and pelagic pathways (e.g. Davenport & Bax 2002; Woodland & Secor 2013; Kopp et al. 2015; Giraldo et al. 2017). Species feeding in the pelagic pathway tend to be have relatively low $\delta^{13}C$ values compared to species feeding on benthic material (e.g. Le Loc'h et al. 2008; Kopp et al. 2015) owing to differences in the $\delta^{13}C$ of primary producers, detritus and associated bacteria remineralising carbon (e.g. France 1995; Bouillon & Boschker 2006; Middelburg 2014). Diettissue fractionation of ^{13}C is often assumed to be small (<1‰), but in practice, for marine fish, fractionation may be higher, with values of 1-2‰ more appropriate (Sweeting, Barry, Barnes, Polunin, & Jennings, 2007)

Sulfur stable isotopes are also expected to provide effective discrimination of marine pelagic and benthic pathways but are only rarely used for this purpose, potentially due to the relatively high cost (Connolly, Guest, Melville, & Oakes, 2004; Barnes & Jennings, 2007; Michener & Kaufman, 2007). Producers predominately deriving sulfur from seawater sulfates such as phytoplankton tend to have δ^{34} S of ca. 19-21‰ whilst producers utilising sedimentary sulfides will have lower δ^{34} S values (Thode, 1991) by as much as 30‰ relative to marine sulfates (Thode, 1991; Connolly *et al.*, 2004;Barros *et al.*, 2010). Sulfur also has the advantage that there is little or no diet-tissue fractionation in δ^{34} S between a fish consumer and diet (0-1‰, δ^{34} S Barnes & Jennings 2007) so errors in assumptions about diet-tissue fractionation will be smaller and they will have a relatively small influence on the estimated contributions of different sources in mixing models (e.g. Parnell *et al.*, 2010), although fractionation may vary more across the entire food chain when differing metabolic pathways are present.

Here, we establish a systematic approach for estimating the contributions of benthic and pelagic pathways to individuals, species and a community. We develop methods that use C and S stable isotope data to estimate the relative contributions of predominantly pelagic food chains supported by phytoplankton consumers (dubbed "pelagic pathways") and benthic food chains supported by detrital pathways ("benthic pathways") to the biomass of 15 species of shelf sea fish in the North Sea. Then we compare the relative contributions of benthic and pelagic pathways as determined from combined analysis of both carbon and sulfur stable isotopes compared to using carbon stables alone. Finally, we assess inter-annual and consumer size-related variation in the contributions of benthic and pelagic pathways to biomass and estimate the proportion of total fish community biomass supported by these pathways.

Materials and methods

Fish sampling and SIA

Fish were caught in an area from 57.5° N - 61.5° N to 1° W - 4° E (mean depth 155 m) in August and/or September from 2002 to 2006 during the North Sea English Bottom Trawl Survey, using a Grande Ouverture Verticale bottom-fished otter trawl net with a 20mm cod-end liner which was towed at approximately four knots for thirty minutes. Fifteen fish species which had the highest rank biomass in the North Sea English Bottom Trawl survey during 2000 and 2001 in the sampling area were caught each year, with an aim to collect up to four individuals from different length classes which spanned the range of total body lengths caught in 2000 and 2001.

Individuals' total body mass was recorded to the nearest 0.1g wet after blotting to remove surface water, or to 1g for larger fishes (typically > 1kg). One to five cm³ of white muscle tissue from the dorsal musculature of each individual was dissected and immediately frozen to -20°C. In the laboratory the frozen stored samples were freeze dried and ground to a homogeneous fine powder using a pestle and mortar. After processing each sample, all equipment was cleaned. Individual samples were transferred to glass vials prior to weighing into tin capsules for SIA.

The carbon stable isotope composition of samples was determined with a Europa Scientific 20-20 IRMS with a Europa Scientific Roboprep-CN preparation module operated by Iso-Analytical Ltd (Crewe, UK). In the preparation module two reference samples of Iso-Analytical Standards IA-R014 (powdered bovine liver), IA-R005 (beet sugar) and IA-R045 and IA-R046 (ammonium sulfate) were placed between every four to six one mg samples of dried fish tissue. The carbon isotope composition of tissue samples was expressed in conventional delta notation (δ^{13} C), relative to the abundance of 13 C in Pee Dee Belemnite. Twenty percent of samples were processed in duplicate for quality control. Standard deviation for samples within batches for a duplicated pair tended to be slightly higher than for replicates of reference material (but <0.25‰) and the 95th percentile of the overall distribution of absolute differences in each duplicated pair was 0.21‰. Experimental precision for all batches of samples was < 0.1‰.

The C:N dry mass ratios of the samples were also determined in the same analysis; when sample C:N ratios were higher than 3.5 (see Jennings & Cogan, (2015) for C:N values), the δ^{13} C values were lipid corrected (Post et al., 2007). Lipid correction used Equation 1 in Sweeting *et al.*, (2006), which produces estimates of δ^{13} C comparable with those based on chemical lipid extraction for fish tissue (Sweeting *et al.*, 2006).

For 34 S analysis, tin capsules containing 2 mg dried fish tissue or reference material plus 4 mg vanadium pentoxide were processed with an automatic sampler coupled to a Europa Scientific 20–20 isotope ratio mass spectrometer operated by Iso-Analytical Limited (Crewe, UK). The reference material used was IA-R036 (Iso-Analytical working standard barium sulfate, δ^{34} S_{V-CDT} +20.74‰) calibrated and traceable to NBS-127 (barium sulfate, δ^{34} S_{V-CDT} +20.3‰). Two capsules containing organic standards, with a composition closer to fish tissue than the inorganic standards, were run after every five tissue samples. The organic standards were IA-R036 and IA-R027 (Iso-Analytical working standard whale baleen, δ^{34} S_{V-CDT} +16.3‰). Analytical precision was good (standard deviation of organic standards < 0.25‰). All results are presented in conventional per mil notation (‰) in relation to the primary 34 S isotope standard V-CDT (troilite of the Canyon Diablo meteorite).

Two source indicator species were selected, to characterise the isotopic composition associated with high dependence on the pelagic or high dependence on the benthic pathway respectively. The affiliation of the remaining 13 species in the community to these pathways was expressed in relation to the dependence of the source indicator species. Each source indicator species was initially identified using a combination of diet and stable isotope data. Diet data were obtained from the Integrated Database and Portal for Fish Stomach Records (DAPSTOM, Pinnegar 2014). All available diet records for the Irish, Celtic, North Sea and English Channel, were used to classify the prey of each fish species in the size-ranges sampled in this study. The prey of each fish species were classified as 'benthic', 'pelagic' or 'unknown' based on position in the water column (Table 1, Supplementary material). Benthic prey were assumed to be all species or groups living on or in the seabed which were more likely to consume material cycled through benthic pathways. Pelagic prey were assumed to be all species or groups living anywhere else in the water column, which were more likely to be supported by food chains based on living phytoplankton. 'Unknown' prey included prey items which were unidentifiable (e.g. partially digested), non-living items (e.g. rock or sand) or items for which the ecology is not sufficiently understood. Mackerel Scomber scombrus Linnaeus, 1758 had consumed the highest proportion of pelagic items by number, while plaice Pleuronectes platessa Linnaeus, 1758 and lemon sole Microstomus kitt (Walbaum, 1792) had consumed the highest proportion of benthic items (Table 1, Supplementary material). Calculations were based on prey number because prey mass data were not available. P. platessa and M.kitt had consumed 99.1 and 99.2% benthic prey respectively, but given that the mean δ^{13} C of *P. platessa* was less negative than the mean for *M.kitt* (-16.9‰ vs -17.9‰), and that this difference would have held even after accounting for the small differences in trophic level between these species (Jennings & van der Molen, 2015), we took plaice as the source indicator species. To test the sensitivities of source indicator species and to assess if results may be affected by the migratory movements of mackerel outside the North Sea, supplementary analyses were conducted using North Sea herring Clupea harengus Linnaeus, 1758 as a source indicator species for pelagic production (Fig. 1, Supplementary material).

 δ^{13} C and δ^{34} S isotopic values of fish consumers were also adjusted to a 'common' trophic level (TL) to account for small trophic level differences which might influence the isotopic values of individuals, independent of any contribution of benthic and pelagic pathways. Median TL estimates (Jennings & van der Molen 2015) were adjusted for trophic fractionation as follows: $\delta^{13}C_{corrected} = \delta^{13}C_{actual} + (-1.74(TL_{actual} - 4.5))$ and $\delta^{34}S_{corrected} = \delta^{34}S_{actual} + (-0.45(TL_{actual} - 4.5))$ where $TL_{actual} - 4.5$ generates an arbitrary common baseline TL that is close to the mean for all sampled individuals and the multipliers represent the mean trophic fractionation of each isotope in teleost fish as reported in Barnes et al. (2007) and Sweeting et al. (2007) respectively. Further analyses were run both with TL corrected and uncorrected isotopic values (TL uncorrected results in Supplementary material).

Biomass estimates were based on numbers-at-length recorded in the International Bottom Trawl Survey in the area 0°-4°E and 56°-62°N in 2006. Catch numbers by species and length per hour of trawling were converted to mean numerical density per ICES statistical rectangle using information on haul meta-data, gear dimensions and gear efficiency (Walker, Maxwell, Le Quesne, & Jennings, 2017). For each species, mean numerical density by length was converted to total numbers-at-length for the whole study area by multiplying the numerical density in each rectangle by the area of the rectangle and summing over all rectangles. Estimates of biomass by species and length were then obtained by converting individual lengths (L, cm) to individual weights (W, g) using the relationship $W=aL^b$, with parameters a and b from Silva et al. (2013), and multiplying individual weights by numbers-at-length.

Mixing models

Two end-member Bayesian mixing models were used to estimate the contribution of the pelagic and benthic pathways to fish biomass. Both models were based on simmr (Parnell, 2016), one using $\delta^{13}C$ and $\delta^{34}S$ as inputs and one using only $\delta^{13}C$.

The model equations were equations 1-4 in Parnell et al. (2013). The source values s_{jk} are assumed to be normally distributed based on the mean and standard deviation of the stable isotopic composition of the two source indicator species (Fig. 2, Supplementary Material). A prior is given for the dietary proportions of each individual i on source k (denoted p_{ik}) based on the centralised log-ratio normal distribution. The residual standard deviation (denoted σ_{ij} for isotope j) was given a uniform prior distribution from 0-100. Trophic fractionation factors or concentration dependencies were not included.

Individuals were grouped by species and 100,000 iterations were run to generate a posterior probability density based on estimated contributions of the pelagic pathway to biomass. The mixing model using $\delta^{13}C$ and $\delta^{34}S$ data was run for the 2006 data and the model using just $\delta^{13}C$ was run for every year from 2002 to 2006. Output comprised estimates of modal and 50% credible intervals of the posterior distribution.

To examine the effects of body size (mass) within species and year on the contribution of the pelagic pathway, the two end-member Bayesian mixing model described above was adapted to include covariates. The model was developed in R using packages R2jags (Su & Yajima, 2015) and plyr (Wickham, 2011). The model was run grouping all species together to examine the overall effect of mass and year. Species were also separated to examine the effect of mass and year on individual species. One million iterations were run for all individuals and 100,000 iterations for individual species. Output comprised estimates of modal and 50% credible intervals of the contribution of the pelagic pathway in centralised log-ratio transformed space, a value for convergence of the model for all parameters (\hat{R}) and an estimate for the amount of variation not explained by the parameters (σ). We only present results for models where the \hat{R} value was close to 1, indicating model convergence has occurred.

Results

Estimates of the relative contributions of pelagic and benthic pathways to fish biomass in 2006, as generated with combined $\delta^{34}S$ and $\delta^{13}C$ (SC) analysis, were more precise than those generated with $\delta^{13}C$ (C) alone (Fig. 1). However, the rank order relative contribution of pelagic pathways, based on relative modal importance of the pelagic pathway, was almost identical (Fig. 1), with a Spearman rank correlation between modes of the credible intervals of 0.96, and 0.90 and 0.93 correlations for the lower and upper bounds of the 50% credible interval. Contributions of the pathways are described as relative because they are expressed in relation to the assumed pelagic contributions of mackerel and plaice, which were taken as 0.99 and 0.01 in the absence of data or a mechanism to define absolute estimates.

The overall relative contributions of pelagic pathways to fish biomass in 2006, based on the relative modal values from the SC and C models and the estimates of fish biomass, were 71% and 59% respectively (Table 1). This result implies that 29 or 41% of biomass was sustained by energy that had cycled through benthic detrital pathways. *C. Harengus* and saithe *Pollachius virens* (Linnaeus, 1758),

the most abundant species in the community, had 86% and 66% pelagic affiliations respectively (Fig. 1, SC estimates).

In 2006, based on both SC and C, there was a marked decrease in pelagic affiliation and an increase in benthic affiliation with body mass for the community as a whole. The larger value of σ for C demonstrated that more variation was unexplained when S data were not included in the analyses (Table 2).

Table 1: Estimated relative contribution of pelagic pathways (modal SC and C estimates) to total biomass (tonnes) in the sampling area (estimated contribution to pelagic pathway by mackerel and plaice assumed to be 0.99 and 0.01 respectively).

Species name	Common name	Authority	Species' biomass (t)	Biomass "pelagic" (t) δ ³⁴ S & δ ¹³ C	Biomass "pelagic" (t) δ ¹³ C
Scomber scombrus	Mackerel	Linnaeus, 1758	324104	320863	320863
Clupea harengus	Herring	Linnaeus, 1758	643212	551282	447761
Trisopterus esmarkii	Norway pout	(Nilsson, 1855)	144739	110586	88489
Pollachius virens	Saithe	(Linnaeus, 1758)	1460296	958157	752536
Limanda limanda	Dab	(Linnaeus, 1758)	62670	40964	36284
Eutrigla gurnardus	Grey gurnard	(Linnaeus, 1758)	99286	60872	53974
Melanogrammus aeglefinus	Haddock	(Linnaeus, 1758)	261692	158852	138752
Hippoglossoides platessoides	Long rough dab	(Fabricius, 1780)	30328	17732	13971
Trachurus trachurus	Scad	(Linnaeus, 1758)	2508	1371	1087
Microstomus kitt	Lemon sole	(Walbaum, 1792)	6830	2575	2014
Amblyraja radiata	Starry ray	(Donovan, 1808)	4892	1679	1373
Merlangius merlangus	Whiting	(Linnaeus, 1758)	90294	28094	24080
Lophius piscatorius	Anglerfish	Linnaeus, 1758	26976	8218	6435
Gadus morhua	Cod	Linnaeus, 1758	26554	7151	6513
Pleuronectes platessa	Plaice	Linnaeus, 1758	7317	73	73
		(Totals)	(3191700)	(2268470)	(189420)

Table 2: Effects of body mass on use of pelagic (p) and benthic (b) pathways in the North Sea during 2006 (mode and 50% credible intervals). Positive values indicate increasing reliance on the relevant pathway. σ is an estimate for the amount of variation not explained by the parameters.

	δ^{13} C & δ^{34} S			$\delta^{13}C$		
	25%	Mode	75%	25%	Mode	75%
Body mass[p]	-1.21	-0.68	-0.17	-1.11	-0.59	-0.07
Body mass[b]	0.17	0.69	1.20	0.15	0.67	1.18
σ	0.90	0.92	0.93	1.11	1.14	1.17

Interannual changes in the relative contribution of pelagic pathways to species' biomass, as determined with δ^{13} C data, were modest (Fig. 2), and correlations between years for the relative modal contributions of pelagic pathways were always >0.57 (Table 3). The Bayesian model with a 'year' covariate reinforced this result, showing only weak year effects (Table 4). The weak year effect detected was negative (mode, -0.04), implying a fall in the relative proportion of biomass supported by pelagic pathways from 2002 to 2006.

Table 3: Correlations, between years, of relative modal contributions of pelagic pathways to species' biomass.

Year	2002	2003	2004	2005	2006
2002	-	0.88	0.65	0.73	0.63
2003		-	0.74	0.76	0.57
2004			-	0.93	0.79
2005				-	0.78
2006					-

Table 4: The effect of body mass and year on the relative contribution of pelagic (p) and benthic (b) pathways to North Sea fish biomass from 2002 to 2006 (mode and 50% credible intervals). Positive values indicate increasing reliance on the relevant pathway.

	25%	Mode	75%
Body mass [p]	-3.49	-2.93	-2.37
Body mass [b]	2.38	2.93	3.48
Year [p]	-0.50	-0.04	0.45
Year [b]	-0.42	0.05	0.54
σ	1.30	1.32	1.33

The effect of body mass on the relative use of the pelagic pathway by the whole fish community was negative and much stronger than the effect of year (Table 4). The high σ indicates that much of the variation in relative use of the pelagic pathway is unexplained by year or mass.

The effects of year and body mass on the relative contribution of the pelagic pathway (Fig. 3) or benthic pathway (Fig. 3, Supplementary Material) to individual species' biomass from 2002-2006 were species dependent. The negative relationship between mass and the relative use of the pelagic pathway was especially strong for whiting *Merlangius merlangus* (Linnaeus, 1758), dab *Limanda limanda* (Linnaeus, 1758), starry ray *Amblyraja radiata* (Donovan, 1808) and *M. kitt*. Conversely, body mass was positively related to the relative use of the pelagic pathway for *C. harengus* and grey gurnard *Eutrigla gurnardus* (Linnaeus, 1758).

Discussion

When attempting to understand the significance of pelagic and benthic food web pathways, our method provides an important complement to diet studies. This is because it characterises the extent to which production is channelled through pelagic or benthic systems as opposed to focusing on the habitat choice of prey or where those prey are consumed. Thus, our results show that even those species feeding on the seabed rely significantly on pelagic production, probably because a proportion of their bottom-living prey are feeding directly on phytoplankton (Vedel, 1998; Lehane & Davenport, 2002; Jumars, Dorgan, & Lindsay, 2015;). Diel vertical migration of zooplankton may also play a significant role in the channelling of pelagic production to benthic consumers (Ringelberg, 2010). Conventional diet studies describing the extent to which consumers feed on prey found in pelagic or benthic environments may not indicate the extent to which consumers are supported by

food chains supplied by phytoplankton grazing or benthic detrital chains. This is because some benthic fauna gain a substantial proportion of their energy from grazing pelagic phytoplankton or zooplankton in shallow shelf seas (Vedel, 1998; Lehane & Davenport, 2002; Jumars, Dorgan, & Lindsay, 2015) and because invertebrates and fish accessible to predators feeding in the water column may feed extensively on benthic detritus and food chains supported by benthic detritus (Mauchline, 1980; Marcus & Marcus, 1998; Jumars, 2007). Nektobenthic organisms, for example, are well recognised as important prey of pelagic fishes such as *C. harengus* (Casini, Cardinale, & Arrhenius, 2004) as well as bottom-dwelling fishes (Mauchline, 1982).

Results implied that the use of the pelagic feeding pathway by *C. harengus* increased with body mass. This was counter to the trend for other species. Previous studies of the trophic role of *C. harengus* based on nitrogen stable isotope analysis have implied that trophic level consistently decreases with body mass, falling by almost one trophic level as mass increases from 30g to 300g (Jennings, D'Oliveira, & Warr, 2007). There are at least two potential drivers of the apparent increase in use of the pelagic pathway and decrease in trophic level. First, interactions between the spatial distributions of base δ^{13} C in the North Sea and age-specific *C. harengus* distributions. Second, real changes in *C. harengus* feeding ecology with age.

In relation to the first driver, Bierman et al., (2010) described the spawning origin of C. harengus caught in the central and northern North Sea in summers of 2004–2007. At the latitudes of our sampling stations, autumn spawning C. harengus (Banks, Buchan and Shetland herring) tended to be most abundant, but winter spawners from the southern North Sea (Downs herring) are also present. Autumn spawners spawn in several inshore areas to the west of the study region and young C. harengus tend to stay in relatively shallow water (<100m) for at least their first year (typically weighing <30g). With age, they move further offshore, predominantly appearing in the study region at an age of 2 or 3 yr (typically 120-140 g). Winter spawners spend their first year in the southern and central North Sea. The spatial distribution of one measure of base δ^{13} C in the North Sea (Barnes, Jennings, & Barry, 2009) varied from -17.6 to -18% in the study region, compared with -17.4 to -17.8% in the inshore areas used by young autumn spawning C. harengus and -16 to -17% in the southern areas used by young winter spawners and adult winter spawners during their migrations. Consequently, autumn spawning C. harengus that have just recruited to the study region may have marginally higher δ^{13} C than older *C. harengus*, but the effect on estimates of the relative proportion of pelagic production and trophic level is likely to be small. For winter spawning C. harengus the effect may be much larger, and given interannual variation in the extent of mixing of autumn and winter spawners in the central North Sea (Bierman et al., 2010), the effects of movement on an apparent increase in use of the pelagic pathway and decrease in trophic level cannot be discounted.

In relation to the effects of real changes in feeding ecology, the increase in the use of the pelagic pathway and decrease in trophic level with body mass in *C. harengus* may reflect an increase in the prevalence of filter feeding with body mass. We are not aware that such a shift has been studied or documented with diet data (Petitgas, 2010), but methods for examining gut contents as used in existing diet studies are unlikely to record phytoplankton feeding effectively. For other clupeid species, where patterns of direct phytoplankton consumption have been a research focus, an increase in phytoplankton feeding and decrease in trophic level with body size results from increased filter feeding and decreased particulate feeding. In sardine *Sardina pilchardus* (Walbaum, 1792), for example, Bode, Carrera, & Lens (2003) demonstrated that δ^{15} N of *S. pilchardus* \geq 18 cm in total length fell with body size. This reduction in δ^{15} N is consistent with a decrease in trophic level (Bode et al., 2007), which would result from increased feeding on phytoplankton by larger fish in cases when

phytoplankton were abundant (Garrido, Marçalo, Zwolinski, & Van der Lingen, 2007). Consumption of phytoplankton by filter feeding *S. pilchardus* may be considerable when large phytoplankton are abundant. For example, chain-forming diatoms may account for 99% of prey ingested by *S. pilchardus* during spring upwelling events off the Portuguese coast (Garrido, 2003). Despite the likely increase in the prevalence of phytoplankton in the diets of filter feeding fish, the absolute trophic levels of *S. pilchardus* (Bode et al., 2017) and *C. harengus* (Jennings & van der Molen, 2015) both imply that zooplankton still contribute substantially to assimilated diet of larger fish. In the future, studies to quantify the role of phytoplankton in ingested and assimilated diet of herring would be valuable. This is because confirmation of reductions in trophic level and increases in direct use of phytoplankton with body size would substantially alter our understanding of the structure and function of pelagic food webs in regions where herring are abundant. These regions include our study sites (Table 1) and the North Sea more widely (Sparholt, 1990).

Our analytical framework provides relative measures of the use of pelagic and benthic pathways. To generate these measures we assumed 99% use of the pelagic and benthic pathways by S. scombrus and P. platessa respectively (the source indicator species). While it is reasonable to assume from diet and stable isotope evidence that S. scombrus primarily use the pelagic pathway and P. platessa primarily use the benthic pathway, their absolute use of either pathway is not known. For these reasons, we always refer to 'relative' use of a pathway. However, we do not expect absolute use of the pathways to differ markedly from our 99% assumptions given that mean depth in the study region was 170 m. Giraldo et al., (2017), for example, used C and N stable isotopes to demonstrate that coupling between pelagic and benthic systems decreases markedly with depth on another part of the northern European continental shelf. At 100m depth, the maximum depth at which they modelled results, the use of the benthic pathway by fishes feeding on the seabed was approximately 85% and the use of the pelagic pathway by pelagic piscivores was approximately 95%. Given P. platessa and S. scombrus are members of these feeding groups, and given that relationships between depth and use of pelagic or benthic pathways were still positive at 100m in the study of Giraldo et al., (2017), it is likely that the assumed 99% use of each pathway at a mean depth of 170m would be within a few percent of the true value (e.g. perhaps within 2% for the pelagic pathway and 10% for the benthic pathway). Thus the errors introduced by estimating pelagic or benthic use for the source indicator species are small in relation to the uncertainties in use of the pathways addressed and expressed in the Bayesian analyses. Since absolute use of the pelagic or benthic pathway by source indicator species is not known, estimates of use of the pelagic or benthic pathway by other species are tied to these. Consequently, conclusions about small changes in the use of pathways by these species through time may also be interpreted as changes in the use of pathways among species and may not indicate absolute changes in reliance on pelagic or benthic pathways at a system level.

In future, it would be desirable to develop the stable isotope methods to provide absolute estimates of pelagic or benthic affiliation in the study region. This would require the identification of species that are specialist feeders on phytoplankton and benthic detritus based food chains respectively and then to sample them with sufficient frequency to account for the short-term isotope dynamics which are evident in smaller individuals with fast turnover times (Wainright & Fry, 1994; Kürten, Painting, Struck, Polunin, & Middelburg, 2013). An alternative is to simulate the seasonal dynamics of δ^{34} S and δ^{13} C in phytoplankton and benthic detritus with models to generate time-integrated estimates of source δ^{34} S and δ^{13} C, as now being attempted for δ^{15} N by biological oceanographers (Somes et al., 2010).

The use of two end-member Bayesian mixing models with both CS and C stable isotope data provided comparable estimates of the relative use of pelagic and benthic pathways by marine fishes in the North Sea, although uncertainty around modal estimates increased when C was used in isolation. The effects of adding S to the mixing model on the precision of the result was unsurprising given the low trophic fractionation of S (which would have minimised any variance in estimates of the relative use of pelagic and benthic pathways introduced by inaccurate estimates of trophic level) and the large and consistent differences between $\delta^{34}S$ of sulfates and sulfides (Thode, 1991; Connolly et al., 2004;Barros et al., 2010). Our conclusion that S led to more precise estimates of source contributions was consistent with the results of Connolly et al., (2004), where a meta-analysis was used to assess the extent to which $\delta^{15}N$, $\delta^{13}C$ and $\delta^{34}S$ distinguished sources in estuarine and marine food webs and affected the precision of estimates of source contributions to consumers. When mixing models included both C and S, the credible intervals around mean estimates of source contributions to consumers were reduced, even though S showed greater within source variability than C.

Relative use of pelagic and benthic pathways varied considerably among species, but more than two-thirds of the biomass in the total fish community was linked to food chains that were ultimately supported primarily by direct phytoplankton grazing rather than production recycled through benthic detrital pathways. Within the community, differences in the relative use of pelagic and benthic pathways by different species were largely consistent from year to year, despite potential fluctuations in planktonic activity, likely reflecting the relatively constrained habitats and diets of most species (Heessen, Daan, & Ellis, 2015) as constrained by morphology and other species' characteristics determined on evolutionary time-scales (Reecht, Rochet, Trenkel, Jennings, & Pinnegar, 2013).

Approximately 30% of total consumer biomass was linked to the benthic pathway, demonstrating that a substantial proportion of fish biomass, and by inference production, in the northern North Sea is supported by production that has passed through transformations on the seabed. All sampled species appear to use both pelagic and benthic pathways to some extent. Accessing both pathways is likely to provide a more resilient feeding strategy than reliance on one or other pathway because detrital-based production cycles are typically dampened and attenuated in relation to cycles in primary production and, together, these out of phase cycles will stabilise temporal variance in total production (Ruardij & Van Raaphorst, 1995; Mussap & Zavatarelli, 2017) and thus food supply to a range of consumers (Darnell, 1961; Kopp et al., 2015). Further, there is strong theoretical evidence to suggest that the presence and use of pathways supported by both primary production and detritus stabilise food webs (McCann et al., 2005) and coupling between these pathways may increase food web resilience to human and environmental perturbations (Blanchard et al., 2011).

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Authors' contributions

J.D.T., S.J., N.P. and A.M. conceived the ideas and designed the methodology; A.P. designed the mixing models used to analyse the data; J.D.T., N.W. and S.J. extracted and analysed the data and J.D.T. led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

Data accessibility

Data used in this study are available from the following sources:

 $\delta^{34}S$ for fish consumers is available from the Dryad Digital Repository:

http://dx.doi.org/10.5061/dryad.j6m2267 (Duffill Telsnig et a. 2018). δ^{13} C and C:N for fish consumers is from Jennings and Cogan (2015). Fish survey length and catch rate data is available from ICES (2016). δ^{13} C scallop data is available from the Dryad Digital Repository:

https://doi.org/10.5061/dryad.sj4fn (Barnes Jennings Barry 2009). Fish stomach contents is available from Pinnegar, J.K. (2014). Trophic level estimates for fish consumers is available from Jennings and van der Molen (2015). Code used to create the bayesian mixing models is available from Zenodo: https://doi.org/10.5281/zenodo.1476659 (Telsnig 2018).

References

- Barnes, C., & Jennings, S. (2007). Effect of temperature, ration, body size and age on sulphur isotope fractionation in fish. *Rapid Communications in Mass Spectrometry*, *21*(8), 1461–1467. doi:10.1002/rcm.2982
- Barnes, C., Jennings, S., & Barry, J. T. (2009). Environmental correlates of large-scale spatial variation in the d13C of marine animals. *Estuarine, Coastal and Shelf Science*, *81*(3), 368–374. doi:10.1016/j.ecss.2008.11.011
- Barnes C, Jennings S, Barry JT (2009) Data from: Environmental correlates of large-scale spatial variation in the δ 13C of marine animals (and related published studies of carbon and nitrogen isotopic baselines). *Dryad Digital Repository*. https://doi.org/10.5061/dryad.sj4fn
- Barnes, C., Sweeting, C. J., Jennings, S., Barry, J. T., & Polunin, N. V. C. (2007). Effect of temperature and ration size on carbon and nitrogen stable isotope trophic fractionation. *Functional Ecology*, 21(2), 356–362. doi:10.1111/j.1365-2435.2006.01224.x
- Barros, N. B., Ostrom, P. H., Stricker, C. A., & Wells, R. S. (2010). Stable isotopes differentiate bottlenose dolphins off west-central Florida. *Marine Mammal Science*, *26*(2), 324–336. doi:10.1111/j.1748-7692.2009.00315.x
- Bierman, S. M., Dickey-Collas, M., van Damme, C. J. G., van Overzee, H. M. J., Pennock-Vos, M. G., Tribuhl, S. V., & Clausen, L. A. W. (2010). Between-year variability in the mixing of North Sea herring spawning components leads to pronounced variation in the composition of the catch. *ICES Journal of Marine Science*, *67*, 885–896.

- Blanchard, J. L., Law, R., Castle, M. D., & Jennings, S. (2011). Coupled energy pathways and the resilience of size-structured food webs. *Theoretical Ecology*, *4*(3), 289–300. doi:10.1007/s12080-010-0078-9
- Bode, A., Alvarez-Ossorio, M. T., Cunha, M. E., Garrido, S., Peleteiro, J. B., Porteiro, C., ... Varela, M. (2007). Stable nitrogen isotope studies of the pelagic food web on the Atlantic shelf of the Iberian Peninsula. *Progress in Oceanography*, 74(2–3), 115–131. doi:10.1016/j.pocean.2007.04.005
- Bode, A., Carrera, P., González-Nuevo, G., Nogueira, E., Riveiro, I., & Santos, M. B. (2017). A trophic index for sardine (Sardina pilchardus) and its relationship to population abundance in the southern Bay of Biscay and adjacent waters of the NE Atlantic. *Progress in Oceanography*. doi:10.1016/j.pocean.2017.08.005
- Bode, A., Carrera, P., & Lens, S. (2003). The pelagic foodweb in the upwelling ecosystem of Galicia (NW Spain) during spring: natural abundance of stable carbon and nitrogen isotopes. *ICES Journal of Marine Science*, 60(1), 11–22. doi:10.1006/jmsc.2002.1326
- Bouillon, S., & Boschker, H. T. S. (2006). Bacterial carbon sources in coastal sediments: a cross-system analysis based on stable isotope data of biomarkers. *Biogeosciences*, *3*(2), 175–185. doi:10.5194/bg-3-175-2006
- Burrel, D. C. (1988). Carbon flow in fjords. Mar. Biol. Ann. Rev., 26, 143–226.
- Carstensen, J., Conley, D. J., & Müller-Karulis, B. (2003). Spatial and temporal resolution of carbon fluxes in a shallow coastal ecosystem, the Kattegat. *Marine Ecology Progress Series*, *252*, 35–50. doi:10.3354/meps252035
- Casini, M., Cardinale, M., & Arrhenius, F. (2004). Feeding preferences of herring (Clupea harengus) and sprat (Sprattus sprattus) in the southern Baltic Sea. *ICES Journal of Marine Science*, *61*(8), 1267–1277. doi:10.1016/j.icesjms.2003.12.011
- Connolly, R. M., Guest, M. A., Melville, A. J., & Oakes, J. M. (2004). Sulfur stable isotopes separate producers in marine food-web analysis. *Oecologia*, *138*(2), 161–167. doi:10.1007/s00442-003-1415-0
- Darnell, R. M. (1961). Trophic Spectrum of an Estuarine Community, Based on Studies of Lake Pontchartrain, Louisiana. *Ecology*, 42(3), 553–568. doi:10.2307/1932242
- Duffill Telsnig, J.I., Jennings, S., Mill, A.C., Walker, N.D., Parnell, A.C., Polunin, N.V.C. (2018) Data from: Estimating contributions of pelagic and benthic pathways to consumer production in coupled marine food webs. *Dryad Digital Repository*. doi:10.5061/dryad.j6m2267
- Davenport, S. R., & Bax, N. J. (2002). A trophic study of a marine ecosystem off southeastern Australia using stable isotopes of carbon and nitrogen. *Canadian Journal of Fisheries and Aquatic Sciences*, *59*(3), 514–530. doi:10.1139/f02-031
- France, R. (1995). Carbon-13 enrichment in benthic compared to planktonic algae:foodweb implications. *Marine Ecology Progress Series*, 124(1–3), 307–312. doi:10.3354/meps124307
- Fry, B. (2006). Stable Isotope Ecology. Springer Berlin. doi:10.1007/s10764-012-9582-7.Fry

- Garrido, S. (2003). Alimentação de Sardina pilchardus (Walbaum,1792) ao largo da costa continental portuguesa e implicações da condição nutricional das fêmeas na qualidade dos oócitos. MS thesis. Instituto de Ciências Biomédicas de Abel Salazar (ICBAS), Porto.
- Garrido, S., Marçalo, A., Zwolinski, J., & Van der Lingen, C. (2007). Laboratory investigations on the effect of prey size and concentration on the feeding behaviour of Sardina pilchardus. *Marine Ecology Progress Series*, *330*, 189–199. doi:10.3354/meps330189
- Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017).

 Depth gradient in the resource use of a fish community from a semi-enclosed sea. *Limnology and Oceanography*, 62(5), 2213–2226. doi:10.1002/lno.10561
- Graf, G. (1992). Benthic-pelagic coupling a benthic view. *Oceanography and Marine Biology, 30,* 149–190.
- Hargrave, B. T. (1973). Coupling Carbon Flow Through Some Pelagic and Benthic Communities. *Journal of the Fisheries Research Board of Canada*, *30*(9), 1317–1326. doi:10.1139/f73-212
- Heessen, H. J. L., Daan, N., & Ellis, J. R. (2015). Fish Atlas of the Celtic Sea, North Sea and Baltic Sea: Based on International Research Vessel Data. KNNV Publishing, The Netherlands.
- ICES (2016) Database of Trawl Surveys (DATRAS), Extraction 20-05-2016. International Bottom Trawl Survey (IBTS) data 2006; http://datras.ices.dk. ICES, Copenhagen.
- Jennings, S., & Cogan, S. M. (2015). Nitrogen and carbon stable isotope variation in northeast Atlantic fishes and squids. *Ecology*, *96*(9), 2568–2568. doi:10.1890/15-0299.1
- Jennings, S., D'Oliveira, J. A. A., & Warr, K. J. (2007). Measurement of body size and abundance in tests of macroecological and food web theory. *Journal of Animal Ecology*, *76*(1), 72–82. doi:10.1111/j.1365-2656.2006.01180.x
- Jennings, S., & van der Molen, J. (2015). Trophic levels of marine consumers from nitrogen stable isotope analysis: estimation and uncertainty. *ICES Journal of Marine Science: Journal Du Conseil*, 72(8), 2289–2300. https://doi.org/10.1093/icesjms/fsv120
- Jumars, P. (2007). Habitat Coupling by Mid-Latitude, Subtidal, Marine Mysids. *Oceanography and Marine Biology*, *45*, 89–138. doi:10.1201/9781420050943.ch3
- Jumars, P. A., Dorgan, K. M., & Lindsay, S. M. (2015). Diet of Worms Emended: An Update of Polychaete Feeding Guilds. *Annual Review of Marine Science*, 7(1), 497–520. doi:10.1146/annurev-marine-010814-020007
- Kiørboe, T., Lundsgaard, C., Olesen, M., & Hansen, J. L. S. (1994). Aggregation and sedimentation processes during a spring phytoplankton bloom: A field experiment to test coagulation theory. *Journal of Marine Research*, *52*(2), 297–323. doi:10.1357/0022240943077145
- Kopp, D., Lefebvre, S., Cachera, M., Villanueva, M. C., & Ernande, B. (2015). Reorganization of a marine trophic network along an inshore—offshore gradient due to stronger pelagic—benthic coupling in coastal areas. *Progress in Oceanography*, 130, 157–171. doi:10.1016/j.pocean.2014.11.001
- Kürten, B., Painting, S. J., Struck, U., Polunin, N. V. C., & Middelburg, J. J. (2013). Tracking seasonal This article is protected by copyright. All rights reserved.

- changes in North Sea zooplankton trophic dynamics using stable isotopes. *Biogeochemistry*, 113(1–3), 167–187. doi:10.1007/s10533-011-9630-y
- Le Loc'h, F., Hily, C., & Grall, J. (2008). Benthic community and food web structure on the continental shelf of the Bay of Biscay (North Eastern Atlantic) revealed by stable isotopes analysis. *Journal of Marine Systems*, 72(1–4), 17–34. doi:10.1016/j.jmarsys.2007.05.011
- Lehane, C., & Davenport, J. (2002). Ingestion of mesozooplankton by three species of bivalve; Mytilus edulis, Cerastoderma edule and Aequipecten opercularis. *Journal of the Marine Biological Association of the UK*, 82(4), S0025315402005957. doi:10.1017/S0025315402005957
- Levinton, J., & Kelaher, B. (2004). Opposing organizing forces of deposit-feeding marine communities. *Journal of Experimental Marine Biology and Ecology*, 300(1–2), 65–82. doi:10.1016/j.jembe.2003.12.008
- Magozzi, S., Yool, A., Vander Zanden, H. B., Wunder, M. B., & Trueman, C. N. (2017). Using ocean models to predict spatial and temporal variation in marine carbon isotopes. *Ecosphere*, 8(5), e01763. doi:10.1002/ecs2.1763
- Marcus, N. H., & Marcus, N. H. (1998). Minireview: The importance of benthic-pelagic coupling and the forgotten role of life cycles in coastal aquatic systems. *Limnology and Oceanography*, *43*(5), 763–768. doi:10.4319/lo.1998.43.5.0763
- Marsay, C. M., Sanders, R. J., Henson, S. A., Pabortsava, K., Achterberg, E. P., & Lampitt, R. S. (2015). Attenuation of sinking particulate organic carbon flux through the mesopelagic ocean. *Proceedings of the National Academy of Sciences*, *112*(4), 1089–1094. doi:10.1073/pnas.1415311112
- Mauchline, J. (1980). The Biology of Mysids and Euphausiids. Advances in Marine Biology, 18, 1-677.
- Mauchline, J. (1982). The predation of mysids by fish of the Rockall Trough, northeastern Atlantic Ocean. *Hydrobiologia*, *93*(1–2), 85–99. doi:10.1007/BF00008102
- McCann, K. S., Rasmussen, J. B., & Umbanhowar, J. (2005). The dynamics of spatially coupled food webs. *Ecology Letters*, *8*(5), 513–523. doi:10.1111/j.1461-0248.2005.00742.x
- Michener, R. H., & Kaufman, L. (2007). Stable Isotope Ratios as Tracers in Marine Food Webs: An Update. In K. Michener & R. Lajtha (Eds.), *Stable Isotopes in Ecology and Environmental Science* (2nd ed., pp. 238–282). Oxford, UK: Blackwell Publishing Ltd. doi:10.1002/9780470691854.ch9
- Middelburg, J. J. (2014). Stable isotopes dissect aquatic food webs from the top to the bottom. *Biogeosciences*, *11*(8), 2357–2371. doi:10.5194/bg-11-2357-2014
- Moore, J. C., Berlow, E. L., Coleman, D. C., Ruiter, P. C., Dong, Q., Hastings, A., ... Wall, D. H. (2004). Detritus, trophic dynamics and biodiversity. *Ecology Letters*, 7(7), 584–600. doi:10.1111/j.1461-0248.2004.00606.x
- Mussap, G., & Zavatarelli, M. (2017). A numerical study of the benthic–pelagic coupling in a shallow shelf sea (Gulf of Trieste). *Regional Studies in Marine Science*, *9*, 24–34. doi:10.1016/j.rsma.2016.11.002
- Noji, T. T., Noji, C. I.-M., & Barthel, K.-G. (1993). Pelagic-benthic coupling during the onset of winter in This article is protected by copyright. All rights reserved.

- a northern Norwegian fjord. Carbon flow and fate of suspended particulate matter. *Marine Ecology Progress Series*, *93*, 89–99.
- Norén, F., Haamer, J., & Lindahl, O. (1999). Changes in the plankton community passing a Mytilus edulis mussel bed. *Marine Ecology Progress Series*, 191, 187–194. doi:10.3354/meps191187
- Parnell, A. C. (2016). simmr. Retrieved from https://cran.r-project.org/web/packages/simmr/index.html
- Parnell, A. C., Inger, R., Bearhop, S., & Jackson, A. L. (2010). Source Partitioning Using Stable Isotopes: Coping with Too Much Variation. *PLoS ONE*, *5*(3), e9672. doi:10.1371/journal.pone.0009672
- Parnell, A. C., Phillips, D. L., Bearhop, S., Semmens, B. X., Ward, E. J., Moore, J. W., ... Inger, R. (2013). Bayesian stable isotope mixing models. *Environmetrics*, 24(6), 387–399. doi:10.1002/env.2221
- Petitgas, P. (Ed. . (2010). Life cycle spatial patterns of small pelagic fish in the Northeast Atlantic. *ICES Cooperative Research Report*, *No. 306*, 93 pp.
- Pinnegar, J. K. (2014). DAPSTOM An Integrated Database & Portal for Fish Stomach Records. Cefas contract report: DP332, C3746, ME1228.
- Pinnegar, J.K. (2014). DAPSTOM An Integrated Database & Portal for Fish Stomach Records. Version 4.7. Centre for Environment, Fisheries & Aquaculture Science, Lowestoft, UK. February 2014, 39pp. https://www.cefas.co.uk/cefas-data-hub/fish-stomach-records/
- Polunin, N. V. C., & Pinnegar, J. K. (2002). *Trophic Ecology and the Strucutre of Marine Food Webs*. (P. J. B. Hart & J. C. Reynolds, Eds.), *Handbook of Fish and Fisheries* (Vol. 1). Blackwell, Oxford.
- Post, D. M., Layman, C. A., Arrington, D. A., Takimoto, G., Quattrochi, J., & Montaña, C. G. (2007). Getting to the fat of the matter: models, methods and assumptions for dealing with lipids in stable isotope analyses. *Oecologia*, *152*(1), 179–189. doi:10.1007/s00442-006-0630-x
- Reecht, Y., Rochet, M.-J., Trenkel, V. M., Jennings, S., & Pinnegar, J. K. (2013). Use of morphological characteristics to define functional groups of predatory fishes in the Celtic Sea. *Journal of Fish Biology*, 83(2), 355–377. doi:10.1111/jfb.12177
- Ringelberg, J. (2010). *Diel Vertical Migration of Zooplankton in Lakes and Oceans*. Dordrecht: Springer Netherlands. doi:10.1007/978-90-481-3093-1
- Ruardij, P., & Van Raaphorst, W. (1995). Benthic nutrient regeneration in the ERSEM ecosystem model of the North Sea. *Netherlands Journal of Sea Research*, *33*(3–4), 453–483. doi:10.1016/0077-7579(95)90057-8
- Scharf, F., Juanes, F., & Rountree, R. A. (2000). Predator size-prey size relationships of marine fish predators: interspecific variation and effects of ontogeny and body size on trophic-niche breadth. *Marine Ecology Progress Series*, 208, 229–248. doi:10.3354/meps208229
- Silva, J. F., Ellis, J. R., & Ayers, R. A. (2013). Length-weight relationships of marine fish collected from around the British Isles. Science Series Technical Report, Cefas Lowestoft (Vol. 150).
- Somes, C. J., Schmittner, A., Galbraith, E. D., Lehmann, M. F., Altabet, M. A., Montoya, J. P., ... Eby, M. (2010). Simulating the global distribution of nitrogen isotopes in the ocean. *Global*
- This article is protected by copyright. All rights reserved.

- Biogeochemical Cycles, 24(4), 1-16. doi:10.1029/2009GB003767
- Sparholt, H. (1990). An estimate of the total biomass of fish in the North Sea. *Journal Du Conseil, Conseil International Pour l'Exploration de La Mer, 46,* 200–210.
- Steele, J. ., & Collie, J. . (2005). Functional diversity and stability of coastal ecosystems. In *The Sea* (Volume 13, pp. 783–817). MA Harvard University Press.
- Su, Y.-S., & Yajima, M. (2015). R2jags: Using R to Run "JAGS." Retrieved from https://cran.r-project.org/package=R2jags
- Suess, E. (1980). Particulate organic carbon flux in the oceans—surface productivity and oxygen utilization. *Nature*, *288*(5788), 260–263. doi:10.1038/288260a0
- Sweeting, C. J., Barry, J., Barnes, C., Polunin, N. V. C., & Jennings, S. (2007). Effects of body size and environment on diet-tissue δ 15N fractionation in fishes. *Journal of Experimental Marine Biology and Ecology*, 340(1), 1–10. doi:10.1016/j.jembe.2006.07.023
- Sweeting, C. J., Barry, J. T., Polunin, N. V. C., & Jennings, S. (2007). Effects of body size and environment on diet-tissue δ13C fractionation in fishes. *Journal of Experimental Marine Biology and Ecology*, 352(1), 165–176. doi:10.1016/j.jembe.2007.07.007
- Sweeting, C. J., Polunin, N. V. C., & Jennings, S. (2006). Effects of chemical lipid extraction and arithmetic lipid correction on stable isotope ratios of fish tissues. *Rapid Communications in Mass Spectrometry*, *20*(4), 595–601. doi:10.1002/rcm.2347
- Telsnig, J (2018). jtelsnig/Duffill-Telsnig-et-al-2018: First release (Version V1.0). *Zenodo*. http://doi.org/10.5281/zenodo.1476659
- Thode, H. (1991). Sulfur isotopes in nature and the environment: an overview. In H. Krouse & V. Grinenko (Eds.), *Stable isotopes: natural and anthropogenic sulphur in the environment* (pp. 1–26). Wiley, Chichester.
- Vedel, A. (1998). Phytoplankton depletion in the benthic boundary layer caused by suspension-feeding Nereis diversicolor (Polychaeta):grazing impact and effect of temperature. *Marine Ecology Progress Series*, 171, 125–132. doi:10.3354/meps171125
- Wainright, S. C., & Fry, B. (1994). Seasonal Variation of the Stable Isotopic Compositions of Coastal Marine Plankton from Woods Hole, Massachusetts and Georges Bank. *Estuaries*, *17*(3), 552. doi:10.2307/1352403
- Walker, N. D., Maxwell, D. L., Le Quesne, W. J. F., & Jennings, S. (2017). Estimating efficiency of survey and commercial trawl gears from comparisons of catch-ratios. *ICES Journal of Marine Science*, 74(5), 1448–1457. doi:10.1093/icesjms/fsw250
- Wickham, H. (2011). The Split-Apply-Combine Strategy for Data Analysis. *Journal of Statistical Software*, 40(1), 1–29. Retrieved from http://www.jstatsoft.org/v40/i01/
- Woodland, R. J., & Secor, D. H. (2013). Benthic-pelagic coupling in a temperate inner continental shelf fish assemblage. *Limnology and Oceanography*, *58*(3), 966–976. doi:10.4319/lo.2013.58.3.0966

